



## Animal, Poultry and Fish Production Research

Available online at <http://zjar.journals.ekb.eg>  
<http://www.journals.zu.edu.eg/journalDisplay.aspx?JournalId=1&queryType=Master>



### EFFICACY OF USING PROBIOTICS CONTAINING *Bacillus subtilis* AND *Bacillus licheniformis* SPORES ON PERFORMANCE AND HEALTH OF HOLSTEIN SUCKLING CALVES

Ahmed E. Shetawy\*, S.M. Bassiony, U.M. Abd El-Moniem and Kh.M. Al-Marakby

Anim. Prod. Dept., Fac. Agric., Zagazig Univ., Egypt

Received: 12/10/2022 ; Accepted: 13/11/2022

**ABSTRACT:** The objective of this study was to evaluate the effect of *Bacillus subtilis* (*BS*) and *Bacillus licheniformis* (*BL*) spores supplementation on performance, general health and some serum constituents in Holstein suckling calves. A total number of 36 newborn pure-bred Holstein calves (5 days old, with an average body weight of  $44.27 \pm 0.37$  and  $42.87 \pm 0.55$  Kg for males and females, respectively) were randomly divided in to three homogeneous groups (12 calves per group, 6 of each sex). For 30 consecutive days, all groups were fed whole milk three times daily. Once a day, 0, 10 and 20 g of the tested probiotics powder dissolved in the whole milk to represent G1 (control), G2 and G3, respectively. Each gram of this powder contained  $2 \times 10^9$  cfu of *BS* +  $2 \times 10^9$  cfu of *BL*. Offering the starter mixture to calves began on the 8<sup>th</sup> day of life. The same parameters were investigated for another 30 days after the end of the probiotics treatment. The obtained results revealed that the overall TDMI value did not affected the first 30 days, but it significantly ( $P < 0.001$ ) increased in post-treatment period in G2 than those in G1 and G3. During the two experiment phases, the ADG of G2 calves was better ( $P < 0.001$ ) than those in the other tested groups. At the end of the trial, the G2 calves were 8.68 Kg heavier than the control calves. Consequently, the weaning age was earlier in G2 than in other tested groups. The incidence of diarrhea and its duration declined in G2 calves compared with G1 and G3 calves. The highest percentage of pneumonia incidence was observed with the calves in G1. The concentrations of serum glucose, total protein, creatinine and activities of ALT and AST did not vary ( $P \geq 0.05$ ) among all tested groups. On the other hand, albumin level were decreased ( $P = 0.02$ ) in G2 and G3, while globulin level was elevated ( $P = 0.02$ ) in G2 compared with those in G1, but these levels stayed within the normal range. Additionally, the triglycerides and cholesterol mean values were significantly ( $P = 0.001$ ) lower in G2 and G3 than in G1. However, serum immunoglobulin G concentrations were significantly ( $P = 0.001$ ) increased in G2 and G3 after 15 days of the probiotics addition compared to that in G1. The economic evaluation showed better return with the low dose of probiotics (G1) than the other groups. Conclusively, supplementing whole milk with *BS* plus *BL* spores had beneficial effects on the performance and health of the suckling Holstein calves.

**Key words:** Probiotics, *Bacillus Subtillis*, *Bacillus licheniformis*, Holstein calves, performance, health.

## INTRODUCTION

The world is becoming densely populated day by day. There is desperate need for some efficient yet solution to supply this population's increasing demand for animal protein source.

Probiotics have a vital role in solving this food production problem and replacing the harmful antibiotic use in farm industries. FAO/WHO (2002) addressed probiotics as live active microbes that offer health values for the host animal when appropriately supplemented.

\* Corresponding author: Tel. :+201272896663  
 E-mail address: shetawyahmed79@gmail.com

Currently, farmers provide probiotic feed supplements to poultry, ruminants and fishes. Probiotics are mostly gram-positive bacteria but there also gram-negative bacteria, yeast and fungi (Arora and Kaur, 2020; Park *et al.*, 2016; Afsharmanesh and Sadaghi, 2014; Zhang *et al.*, 2020; Zhang and Kim, 2014; Bai *et al.*, 2013).

Probiotics exert their effectiveness through diverse mechanisms. Probiotics inhibit and control pathogens along with improving the functioning and production capacity of animals (Maas *et al.*, 2021; Van Zyl *et al.*, 2020; Layus, 2020; Chen *et al.*, 2020; Sharma *et al.*, 2018; Mookiah *et al.*, 2014).

Probiotics can enhance milk production (Ma *et al.*, 2020), digestibility (Boyd *et al.*, 2011), immune system (Signorini *et al.*, 2012), improve dietary intake in newborn calves (Muya *et al.*, 2015) and promote the viability and balance of rumen microorganisms (Chen *et al.*, 2020).

Considering the limited and scarce research on using probiotics in newborn animals, more insight into their efficacy and mode of action is needed. The objective of this study was to evaluate the effect of probiotics containing  $2 \times 10^9$  cfu of *Bacillus subtilis* (BS) and  $2 \times 10^9$  cfu of *Bacillus Licheniformis* (BL) on growth performance and health status of Holstein suckling calves.

## MATERIAL AND METHODS

The trial was conducted at the Global dairy farm (29°24'26"N30°52'00"E), Sinnuris, Al-Fayoum, Egypt, during the year 2020 (March to May).

### Animals and Management

A growth experiment was performed on thirty-six neonatal pure-bred Holstein calves weighing  $44.27 \pm 0.37$  and  $42.87 \pm 0.55$  Kg for males and females, respectively. The trial started from the 5<sup>th</sup> day of age and continued till reaching the weaning weight (115 Kg). The animals were randomly divided into three similar groups (12 calves per group, 6 of each sex). Each calf was individually placed in a suckling box (2×1×1 meters) on a sandy bed under shading. All calves were healthy and kept under the same managerial procedures.

### Experimental Design and Treatments

The calves received the whole milk supplemented with 0, 10 and 20 g of the probiotics as a single daily dose to represent G1 (control), G2 and G3, respectively. These probiotics are a formula in a powder form prepared especially for this research in Agrivit Company for manufacturing feed additives, Cairo, Egypt. Each gram of this powder contained  $2 \times 10^9$  cfu of BS and  $2 \times 10^9$  cfu of BL.

Before the evening meal (from 1 to 3 p.m.), the probiotics powder was dissolved well in the suckling bucket and offered. The treatment lasted for 30 days after the colostrum period. Animals always have free access to clean water.

### Feeding Regime

Within an hour of birth, all calves received the first meal (4 liters of colostrum, from their dams) by a stomach tube. Thereafter, calves were fed the transition milk till the end of the 4<sup>th</sup> day by using suckling buckets provided with nipples. Then, the treatment started and all groups were fed the whole milk three times daily in the same technique. Calves obtained standard quantities milk of according to the suckling routine followed on the farm. The offered quantities of whole milk from 1<sup>st</sup> week of life up to 12 weeks increased gradually with the age (5 – 12 Kg / day). This routine is named the accelerated growth program for milk-fed calves and uses the modifications of NRC (2001) equations illustrated by Cornell-Illinois (Van Amburgh and Drackley, 2005).

Calves were fed starter mixture (SM) *ad libitum* in buckets from the beginning of the 2<sup>nd</sup> week of life up to the end of the experimental period. The SM consisted of 90% a mash concentrate feed mixture plus 10% wheat straw. The formulation and proximate chemical analysis of the SM are displayed in Table 1.

### Performance Parameters

Live body weight of calves was estimated by measuring the chest circumference using the standardized weight tape at birth, 15, 30, 45, 60 and the weaning day. The average daily gain (ADG) was calculated by dividing the live weight gain (g) by the growth period length (day).

**Table 1. Formulation and proximate chemical analysis of the starter mixture and milk (on DM basis)**

Items	Kg / Ton
Ingredients of the starter:	
Yellow corn	430
Soya bean meal (46%)	350
Wheat bran	100
Salt	10
Limestone	5
Minerals mixture	3
Vitamins mixture	1.5
Anti-toxin	0.5
Wheat straw	100
Nutrients content:	(%)
Starter mixture:	
Dry matter	90.20
Organic matter	92.37
Crude protein	21.1
Ether extract	4.35
Crude fiber	16.17
Nitrogen free extract	50.75
Ash	7.63
Acid insoluble ash	2.44
GE Mcal/Kg DM	4.87
DE Mcal/Kg DM	3.59
ME Mcal/Kg DM	3.19
Whole Milk:	
Dry matter	12.80
Crude protein	3.35
Fat	3.89
Lactose	4.81
Ash	0.75
GE Mcal/Kg	0.739
DE Mcal/Kg	0.584
ME Mcal/Kg	0.560

Gross energy (GE), digestible energy (DE), and metabolizable energy (ME) of the starter mixture and the whole milk (Mcal/Kg) were calculated from as described below:

GE of the starter mixture =  $(CP \% \times 0.057) + (EE \% \times 0.094) + (\text{Total carbohydrate} \% \times 0.0415)$

DE of the starter mixture =  $GE \times 0.82$

ME of the starter mixture =  $(DE \times 1.01) - 0.45 + (0.0046 \times \text{Fat} \% - 3)$

GE of Milk =  $(CP \% \times 0.057) + (\text{Fat} \% \times 0.092) + (\text{Lactose} \% \times 0.0395)$

DE of Milk =  $GE \times 0.97$

ME of Milk =  $DE \times 0.96$ .

The dry feed intake was recorded daily from the 8<sup>th</sup> day of life till the weaning by subtracting amounts of the residual from the offered. Feed conversion ratio (FCR) was estimated by dividing the total dry matter intake of both milk and the SM (Kg) by the live weight gain (Kg).

Protein and energy utilization parameters were estimated as follows:

Protein content of gain (PCG) =  $30 \times 6.25 \times \text{ADG (Kg)}$ , according to **NRC (2001)**.

Efficiency of crude protein utilization (ECPU %) =  $(\text{PCG}/\text{CPI}) \times 100$ , where, CPI is the crude protein intake

Energy conversion ratio (ECR) =  $\text{ADG gain}/\text{MEI}$ , where MEI is the metabolizable energy intake.

### Disease Incidence

Observations of diarrhea incidence (DI), diarrhea duration (DD) and pneumonia incidence (PI) were detailed as daily notes. Feces were scored according to **Renaud *et al.* (2020)**. Where: 0, 1, 2, and 3 defined as normal, soft, fluid, and aqueous types of feces, respectively, and the score of  $\geq 2$  indicated the presence of diarrhea. The diarrhea duration (scores 2 or 3) is the total number of disease days. However, the percentage of diarrhea incidence is the number of calves in diarrhea divided by the total number of calves in each group.

### Blood Sampling

The first blood sample were collected before first meal of colostrum and the other blood sample were collected after the meal of the morning by 3 hours on day 30<sup>th</sup> and 45<sup>th</sup> from the treatment start. Samples were collected via the jugular vein by using a sterile syringe and then transferred to sterile tubes. The collected samples were centrifuged at 1006 g for 10 minutes. Sera were preserved at -18°C until tested. Commercial kits (DiaSys, Diagnostic Systems GmbH, Germany) were used to assess liver functions, kidney functions, metabolites and immune response parameters.

### Analytical Procedures

#### Proximate chemical analysis

Nutrient contents of the milk and SM samples were determined according to **AOAC (1990)**

while, Nitrogen free extract in the SM was obtained by the difference.

### Blood biochemical and immunological parameters

Levels of serum total protein and albumin were determined by the methods of **Bakker and Mücke (2007)**. While, serum globulin concentration was calculated as follow  $\text{Globulin (g/dL)} = \text{Total protein (g/dL)} - \text{Albumin (g/dL)}$ . Activities of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were determined according to the methods of **Thomas (1998)**. Levels of serum glucose and creatinine were estimated by methods described by **Young *et al.*, (2000)**. Serum triglycerides and cholesterol levels were measured according to **Rifai *et al.* (1999)**. Immunoglobulins IgG was quantitatively determined by Sandwich Enzyme Linked Immunosorbent Assay (ELISA) according to **Engvall and Perlmann (1971)**.

### Economic Evaluation

The economic evaluation was gauged in line with the prevailing market prices. The Relative profit (RP) was computed as follows:

$\text{RP} = (\text{profit of each tested diet}/\text{profit of the control diet}) \times 100$

### Statistical Analysis

Data handling and statistical analysis was carried out at the Dept. of animal production, Faculty of Agriculture, Zagazig University. Analysis was done using SPSS/PCT, (Statistical Package for Social Sciences version 22.0) (IBM Corp., Armonk, NY, USA) software. Results were reported in means  $\pm$  SEM (Standard Error of Mean). The value of  $P < 0.05$  was used to indicate statistical significance. The statistical method was ANOVA test (one way analysis of variance) to test the differences in control and probiotics groups. The Duncan multiple range tests are also used (**Duncan, 1955**).

## RESULTS AND DISCUSSION

The current work was achieved to investigate the impacts of two doses of the probiotics (*BS* and *BL*) on the growth performance, health and economical feed efficiency of Holstein suckling Holstein calves.

## Effect of Probiotics Supplements on Growth Performance

The effect of probiotics supplementation on averages of total dry matter intake (TDMI), live body weight (LBW), daily gain (ADG) and feed conversion ratio (FCR) all over the environmental period are introduced in Table 2.

The probiotics addition had no significant impact on the TDMI values except G2 which significantly ( $P<0.05$ ) improved in the post treatment period compared with the other groups (G1 and G3). However the averages of LBW, ADG and FCR were positively ( $P<0.05$ ) influenced by the lower dose (10 g/day) of probiotics (Table 2). In contrast the higher dose (20 g/day) of probiotics addition (G3) negatively affected the average of LBW, ADG and FCR during the treatment period and significantly ( $P<0.05$ ) improved in the post treatment period, and showed no significant differences between G3 and G1 (83.16 vs. 82.24, respectively) at the end of the trial.

The probiotics as additive in G2 enhanced the overall means of LBW, ADG and FCR by 5.16, 18.14 and 15.05%, respectively, in the treatment period and 10.55, 13.27 and 9.45% in the post treatment period, respectively, compared with the control group (Table 2). At the end of the trial the supplemented calves with lower dose were 8.68Kg heavier than the control and 7.76 Kg than the calves received the higher dose.

The performance improvement of calves fed diet supplemented with 10 g probiotics (G2) positively reflected on the weaning age (Fig. 1) than the control and G3 (78.33 vs. 84.92 and 86.08 days, respectively).

The calf sex did not significantly affect the averages of TDMI and LBW. Conversely, males had superior ( $P<0.05$ ) values of ADG and FCR compared with females (Table 2). The interaction effect between the probiotics and sex did not significantly affect the tested growth parameters through the experimental period.

The results are in agreement with **Smock et al. (2020)** who noted an improvement in ABG and LBW during the initial 56-day feed-lot receiving phase when *B. subtilis* PB6 was supplemented to high stressed feed steers. Also,

**Mousa and Marwan (2019)** found that buffalo calves at 15<sup>th</sup> day old, showed insignificantly greater body weight when supplemented group fed *Bacillus spp.* than the unsupplemented group. However the supplemented group showed significantly increase in body weight at 30<sup>th</sup> day than the control one. Similar results were recorded by **Khalifa et al. (2016)** in lambs and **Kochewad et al. (2009)** in growing kids. The researchers discussed the improving DBG of animals fed probiotics may be a result to favorable growth of useful bacteria which colonized in intestine more quickly the pathogenic bacteria.

Also, **Liao et al. (2010)** and **Kowalski et al. (2009)** showed that bacterial probiotics supplementation numerically improved DM intake and feed conversion ratio in suckling and post-partum period compared with the control. The significant increase may reflect higher ruminal fermentation in the treated groups than in control. However, **Aikman et al. (2011)** observed no difference in DM intake between the control and treated cows fed two TMR's differing in level of concentrate and supplemented with direct-fed-microbial (DFM).

On the other hand, **Mostafa et al. (2014)** found insignificant effect of dietary supplementation of two probiotics on live body weight of cow during pre-partum, calving and post-partum.

The result of feed efficiency, calculated as consumption of metabolizable energy intake (MEI) and crude protein intake (CPI) (Table 3) showed significant ( $P<0.05$ ) improved the efficiency of crude protein utilization (ECPU) and energy conversion ratio (ECR) with the lower dose(G2) compared with control (G1) and the higher dose (G3) which reflected on the better final live weight of calves. These results are in agreement with **Kowalski et al. (2009)** who found that the supplement calves with probiotics were significantly heavier as a result of improving the feed efficiency of consumed ME and CP.

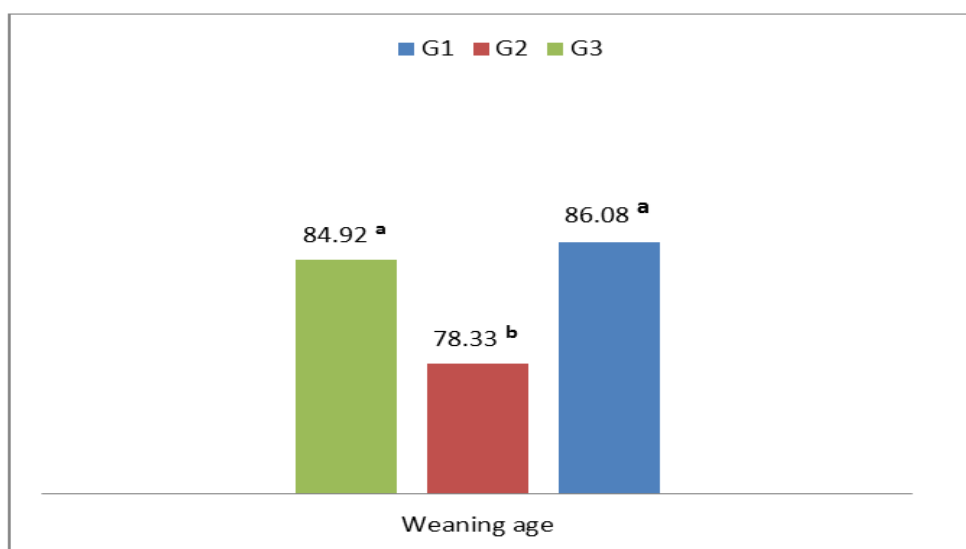
## Effect of Probiotics Supplements on Blood Constituents

Table 4 presents the results of serum biochemical parameters in Holstein calves that suckled milk supplemented with spores of *Bs* and *Bl*.

**Table 2. Effects of probiotics, calves sex and their interaction on growth performance (Mean  $\pm$  SE) during the treatment period and after treatment period**

Items	Initial weight Kg	During treatment period				After treatment period			
		DMI Kg/day	30 <sup>th</sup> day weight Kg	ADG g/day	FCR	DMI Kg/day	60 <sup>th</sup> day weight Kg	ADG g/day	FCR
<b>Groups</b>									
G <sub>1</sub>	43.08 $\pm$ 0.66	1.03 $\pm$ 0.01	58.24 <sup>b</sup> $\pm$ 0.64	498.33 <sup>b</sup> $\pm$ 12.66	2.06 <sup>b</sup> $\pm$ 0.01	1.72 <sup>b</sup> $\pm$ 0.01	82.24 <sup>b</sup> $\pm$ 0.60	866.67 <sup>c</sup> $\pm$ 16.92	2.01 <sup>a</sup> $\pm$ 0.04
G <sub>2</sub>	43.67 $\pm$ 0.31	1.04 $\pm$ 0.01	61.25 <sup>a</sup> $\pm$ 0.30	588.75 <sup>a</sup> $\pm$ 11.80	1.75 <sup>c</sup> $\pm$ 0.04	1.78 <sup>a</sup> $\pm$ 0.01	90.92 <sup>a</sup> $\pm$ 0.36	981.67 <sup>a</sup> $\pm$ 11.13	1.82 <sup>b</sup> $\pm$ 0.02
G <sub>3</sub>	43.54 $\pm$ 0.87	1.03 $\pm$ 0.01	55.39 <sup>c</sup> $\pm$ 0.70	396.67 <sup>c</sup> $\pm$ 18.01	2.68 <sup>a</sup> $\pm$ 0.13	1.72 <sup>b</sup> $\pm$ 0.01	83.16 <sup>b</sup> $\pm$ 0.22	930.56 <sup>b</sup> $\pm$ 21.89	1.89 <sup>b</sup> $\pm$ 0.05
<b>P value</b>	1.00	0.43	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001	<0.001
<b>Sex</b>									
<b>Males</b>	44.27 $\pm$ 0.37	1.03 $\pm$ 0.01	58.60 $\pm$ 0.79	474.91 <sup>b</sup> $\pm$ 22.64	2.27 <sup>a</sup> $\pm$ 0.12	1.74 $\pm$ 0.01	86.60 $\pm$ 0.94	918.52 $\pm$ 18.52	1.93 $\pm$ 0.04
<b>Females</b>	42.87 $\pm$ 0.55	1.03 $\pm$ 0.01	57.96 $\pm$ 0.68	514.26 <sup>a</sup> $\pm$ 20.87	2.06 <sup>b</sup> $\pm$ 0.10	1.74 $\pm$ 0.01	85.57 $\pm$ 0.80	934.07 $\pm$ 17.14	1.89 $\pm$ 0.04
<b>P value</b>	0.07	0.53	0.73	0.02	0.02	0.94	0.82	0.44	0.40
<b>Groups <math>\times</math> Sex</b>									
<b>G<sub>1</sub> <math>\times</math> Males</b>	45.25 $\pm$ 0.85	1.04 $\pm$ 0.01	59.75 $\pm$ 1.11	485.83 $\pm$ 20.38	2.15 $\pm$ 0.09	1.73 $\pm$ 0.01	85.00 $\pm$ 1.08	844.44 $\pm$ 28.11	2.07 $\pm$ 0.06
<b>G<sub>2</sub> <math>\times</math> Males</b>	43.67 $\pm$ 0.56	1.03 $\pm$ 0.01	60.84 $\pm$ 0.40	572.22 $\pm$ 18.09	1.79 $\pm$ 0.06	1.78 $\pm$ 0.01	90.50 $\pm$ 0.56	988.89 $\pm$ 7.03	1.81 $\pm$ 0.02
<b>G<sub>3</sub> <math>\times</math> Males</b>	44.20 $\pm$ 0.49	1.03 $\pm$ 0.01	55.00 $\pm$ 0.71	366.67 $\pm$ 14.91	2.88 $\pm$ 0.16	1.72 $\pm$ 0.02	83.20 $\pm$ 0.37	922.22 $\pm$ 23.83	1.90 $\pm$ 0.06
<b>G<sub>1</sub> <math>\times</math> Females</b>	42.11 $\pm$ 0.68	1.01 $\pm$ 0.01	57.56 $\pm$ 0.71	510.83 $\pm$ 15.06	1.98 $\pm$ 0.06	1.72 $\pm$ 0.01	83.89 $\pm$ 0.73	888.89 $\pm$ 16.48	1.94 $\pm$ 0.04
<b>G<sub>2</sub> <math>\times</math> Females</b>	43.67 $\pm$ 0.77	1.04 $\pm$ 0.01	61.67 $\pm$ 0.42	605.28 $\pm$ 13.27	1.72 $\pm$ 0.04	1.78 $\pm$ 0.01	91.34 $\pm$ 0.42	974.44 $\pm$ 21.79	1.83 $\pm$ 0.04
<b>G<sub>3</sub> <math>\times</math> Females</b>	43.13 $\pm$ 1.39	1.03 $\pm$ 0.01	55.63 $\pm$ 1.08	426.67 $\pm$ 29.08	2.48 $\pm$ 0.18	1.72 $\pm$ 0.02	83.13 $\pm$ 0.30	938.89 $\pm$ 38.88	1.89 $\pm$ 0.09
<b>P value</b>	0.26	0.05	0.16	0.64	0.32	0.78	0.32	0.50	0.36

G1(control): calves suckled whole milk without additives, G2: calves suckled whole milk supplemented with 10 g of probiotics ( $2 \times 10^9$  CFU of *Bs* +  $2 \times 10^9$  CFU of *Bl*), G3: calves suckled whole milk supplemented with 20 g of the same probiotics. a, b and c: Means in the same column with different letters are significantly ( $P < 0.05$ ) differ.



**Fig. 1. Effects of probiotics supplementation on the weaning age in the suckling Holstein calves. a and b: Means with different letters are significantly ( $P < 0.05$ ) differ**

**Table 3. Effects of probiotics, calves sex and their interaction on protein and energy utilization (Mean ± SE) during treatment and a month post-treatment**

Items	During of treatment					
	CPI g/dad	MEI Mcal/day	ADG g/day	PCG g	ECPU %	ECR g/Mcal
<b>Groups</b>						
G <sub>1</sub>	262.10 ±1.12	4.78±0.02	498.33 <sup>b</sup> ±12.66	93.44 <sup>b</sup> ±2.37	35.87 <sup>b</sup> ±0.86	105.15 <sup>b</sup> ±2.50
G <sub>2</sub>	264.79±0.94	4.83±0.02	588.75 <sup>a</sup> ±11.80	110.39 <sup>a</sup> ±2.21	41.72 <sup>a</sup> ±0.76	122.34 <sup>a</sup> ±2.24
G <sub>3</sub>	263.73±0.75	4.81±0.01	396.67 <sup>c</sup> ±18.01	74.38 <sup>c</sup> ±3.38	28.33 <sup>c</sup> ±1.37	83.08 <sup>c</sup> ±4.04
<b>P value</b>	0.11	0.11	<0.001	<0.001	<0.001	<0.001
<b>Sex</b>						
Males	263.70±0.71	4.81±0.01	474.91 <sup>b</sup> ±22.64	89.05 <sup>b</sup> ±4.24	33.87 <sup>b</sup> ±1.62	99.31 <sup>b</sup> ±4.77
Females	263.38±0.89	4.80±0.02	514.26 <sup>a</sup> ±20.87	96.42 <sup>a</sup> ±3.91	36.75 <sup>a</sup> ±1.43	107.74 <sup>a</sup> ±4.18
<b>P value</b>	0.75	0.75	0.02	0.02	0.02	0.02
<b>Groups × Sex</b>						
G <sub>1</sub> × Males	264.35±1.38	4.82±0.02	485.83±20.38	91.09±3.82	34.74±1.27	101.92±3.72
G <sub>2</sub> × Males	263.47±1.57	4.81±0.03	572.22±18.09	107.29±3.39	40.89±1.19	119.90±3.48
G <sub>3</sub> × Males	263.29±0.79	4.80±0.01	366.67±14.91	68.75±2.79	25.98±1.18	76.11±3.49
G <sub>1</sub> × Females	259.86±1.27	4.74±0.02	510.83±15.06	95.78±2.82	37.01±1.07	108.39±3.07
G <sub>2</sub> × Females	266.10±0.84	4.85±0.01	605.28±13.27	113.49±2.49	42.55±0.92	124.78±2.76
G <sub>3</sub> × Females	264.17±1.33	4.82±0.02	426.67±29.08	80.00±3.38	30.68±2.16	90.05±6.35
<b>P value</b>	0.05	0.05	0.64	0.64	0.50	0.50
	Post-treatment					
	CPI g/d	MEI Mcal/day	ADG g/day	PCG g	ECPU %	ECR g/Mcal
<b>Groups</b>						
G <sub>1</sub>	425.34 <sup>b</sup> ±0.89	8.03 <sup>b</sup> ±0.01	866.67 <sup>c</sup> ±16.92	162.50 <sup>c</sup> ±3.17	38.26 <sup>b</sup> ±0.74	108.10 <sup>b</sup> ±2.10
G <sub>2</sub>	437.47 <sup>a</sup> ±0.92	8.24 <sup>a</sup> ±0.02	981.67 <sup>a</sup> ±11.13	184.06 <sup>a</sup> ±2.09	42.07 <sup>a</sup> ±0.47	119.19 <sup>a</sup> ±1.35
G <sub>3</sub>	425.38 <sup>b</sup> ±2.25	8.03 <sup>b</sup> ±0.04	930.56 <sup>b</sup> ±21.89	174.48 <sup>b</sup> ±4.10	40.78 <sup>a</sup> ±1.03	115.18 <sup>a</sup> ±2.89
<b>P value</b>	<0.001	<0.001	<0.001	<0.001	0.01	0.01
<b>Sex</b>						
Males	429.47±1.77	8.10±0.03	918.52±18.52	172.22±3.47	39.97±0.74	113.04±2.11
Females	429.32±1.89	8.10±0.03	934.07±17.14	175.14±3.21	40.76±0.72	115.28±2.05
<b>P value</b>	0.94	0.94	0.44	0.45	0.39	0.39
<b>Groups × Sex</b>						
G <sub>1</sub> × Males	426.30±1.51	8.05±0.03	844.44±28.11	158.33±5.27	37.11±1.08	104.88±3.07
G <sub>2</sub> × Males	436.85±1.60	8.23±0.03	988.89±7.03	185.42±1.32	42.34±0.43	119.93±1.20
G <sub>3</sub> × Males	425.25±3.27	8.03±0.06	922.22±23.83	172.92±4.46	40.47±1.20	114.30±3.36
G <sub>1</sub> × Females	424.37±0.92	8.02±0.02	888.89±16.48	166.67±3.09	39.41±0.86	111.32±2.43
G <sub>2</sub> × Females	438.08±0.99	8.25±0.02	974.44±21.79	182.71±4.09	41.80±0.88	118.45±2.52
G <sub>3</sub> × Females	425.51±3.39	8.04±0.06	938.89±38.88	176.04±7.29	41.09±1.78	116.06±5.02
<b>P value</b>	0.76	0.76	0.50	0.50	0.46	0.46

G1(control): calves suckled whole milk without additives, G2: calves suckled whole milk supplemented with 10 g of probiotics ( $2 \times 10^9$  CFU/g of *Bs* +  $2 \times 10^9$  CFU/g of *Bl*), G3: calves suckled whole milk supplemented with 20 g of the same probiotics. CPI: crude protein content intake, MEI: metabolizable energy intake, ADG: average daily gain, PCG: protein content of gain =  $30 \times 6.25 \times \text{ADG}$  (Kg) according to NRC (2001), ECPU %: efficiency of crude protein utilization= (PCG/CPI)  $\times 100$ , ECR: energy conversion ratio= ADG gain/MEI. a, b and c: Means in the same column with different letters are significantly ( $P < 0.05$ ) differ.

**Table 4. Effects of probiotics supplementation on the serum biochemical parameters (Mean  $\pm$  SE) in the sulking Holstein calves**

Items		Groups			P value
		G <sub>1</sub>	G <sub>2</sub>	G <sub>3</sub>	
Glucose	mg/dL				
	Day 1	117.32 $\pm$ 5.52	116.66 $\pm$ 6.20	119.32 $\pm$ 6.04	0.46
	Day 30	94.00 $\pm$ 2.21	92.00 $\pm$ 4.57	93.50 $\pm$ 5.28	0.06
Total protein	g/dL				
	Day 1	4.72 $\pm$ 0.05	4.62 $\pm$ 0.05	4.73 $\pm$ 0.08	0.45
	Day 30	6.69 $\pm$ 0.21	6.89 $\pm$ 0.05	6.50 $\pm$ 0.09	0.21
Albumin	g/dL				
	Day 1	2.67 $\pm$ 0.04	2.58 $\pm$ 0.05	2.68 $\pm$ 0.08	0.46
	Day 30	3.39 <sup>a</sup> $\pm$ 0.17	2.92 <sup>b</sup> $\pm$ 0.04	3.11 <sup>b</sup> $\pm$ 0.02	0.02
Globulin	g/dL				
	Day 1	2.05 $\pm$ 0.01	2.04 $\pm$ 0.01	2.05 $\pm$ 0.01	0.72
	Day 30	3.30 <sup>b</sup> $\pm$ 0.15	3.97 <sup>a</sup> $\pm$ 0.01	3.39 <sup>b</sup> $\pm$ 0.08	0.01
ALT	IU/L				
	Day 1	7.93 $\pm$ 0.35	8.00 $\pm$ 0.57	8.13 $\pm$ 0.29	0.95
	Day 30	9.46 $\pm$ 0.29	7.94 $\pm$ 0.02	8.00 $\pm$ 0.57	0.05
AST	IU/L				
	Day 1	53.33 $\pm$ 1.45	52.66 $\pm$ 1.45	53.00 $\pm$ 0.57	0.93
	Day 30	48.86 $\pm$ 0.94	51.96 $\pm$ 2.26	49.00 $\pm$ 0.57	0.05
Triglycerides	mg/dL				
	Day 1	39.00 $\pm$ 0.57	39.10 $\pm$ 0.37	38.90 $\pm$ 0.55	0.96
	Day 30	59.66 <sup>a</sup> $\pm$ 0.88	37.00 <sup>c</sup> $\pm$ 0.57	48.00 <sup>b</sup> $\pm$ 0.57	0.001
Cholesterol	mg/dL				
	Day 1	42.53 $\pm$ 1.44	43.03 $\pm$ 0.75	42.53 $\pm$ 0.29	0.91
	Day 30	102.66 <sup>a</sup> $\pm$ 1.45	95.43 <sup>b</sup> $\pm$ 0.26	97.00 <sup>b</sup> $\pm$ 0.57	0.001
Creatinine	mg/dL				
	Day 1	3.25 $\pm$ 0.02	3.25 $\pm$ 0.01	3.35 $\pm$ 0.12	0.58
	Day 30	0.90 $\pm$ 0.08	0.96 $\pm$ 0.03	0.95 $\pm$ 0.05	0.11
IgG	mg/dL				
	Day 45 *	1104.33 <sup>c</sup> $\pm$ 2.33	1169.33 <sup>b</sup> $\pm$ 2.60	1241.66 <sup>a</sup> $\pm$ 7.26	0.001

G1(control): calves suckled whole milk without additives, G2: calves suckled whole milk supplemented with 10 g of probiotics ( $2 \times 10^9$  CFU/g of *Bs* +  $2 \times 10^9$  CFU/g of *Bl*), G3: calves suckled whole milk supplemented with 20 g of the same probiotics. \*: 15 days post-treatment. a, b and c: Means in the same column with different letters are significantly ( $P < 0.05$ ) differ. ALT: Alanine aminotransferase, AST: Aspartate aminotransferase, IgG: Immunoglobulin G.

There were no significant variations ( $P \geq 0.05$ ) in all of serum biochemistry measurements among all treatments on the 1<sup>st</sup> day of the study.

On the 30<sup>th</sup> day of probiotics treatment, concentrations (conc.) of serum glucose and total protein, creatinine and activities of ALT and AST did not alter ( $P \geq 0.05$ ). On the other hand, values of albumin were decreased ( $P = 0.02$ ) by 13.86 and 8.26% in G2 and G3, respectively, while the conc. of globulin was elevated ( $P = 0.02$ ) by 20.30% in G2 compared with those in G1. Additionally, the triglycerides

and cholesterol mean values were significantly ( $P = 0.001$ ) lower in G2 and G3 than those in G1 by 37.98 and 19.54% and 7.04 and 5.51%, respectively. However, 15 days after the end of probiotics treatment, serum immunoglobulin G (IgG) conc. were significantly ( $P = 0.001$ ) increased with calves that received the probiotics treatments (G2 and G3) earlier compared to that in G1.

Many authors agreed with our finding concerning the insignificant impact of probiotics treatment on glucose and/or total protein conc. in Holstein calves (Riddell *et al.*, 2010; Noori



*et al.*, 2016; Fouladgar *et al.*, 2016; Le *et al.*, 2016; Seifzadeh *et al.*, 2017). The insignificant effect of the probiotics treatment on creatinine values is similar to those obtained by Talha *et al.* (2009), Le *et al.* (2016) and Mousa *et al.* (2019).

Despite the significant decrease in albumin conc. with the addition of probiotics to milk, its mean values remained within the normal range (from 2.0 to 2.7 g/dL) which was revealed by Hussein *et al.* (2020) for the suckling Holstein calves. The remarkable elevation of globulin conc. as a result of probiotics inclusion in G2 concurred with that observed by Talha *et al.* (2009) in buffalo calves fed milk supplemented with probiotics. Nevertheless, the globulin level in G2 is still within the normal range (from 3.0 to 4.6 g/dL) published by Hussein *et al.* (2020) on the suckling Holstein calves.

Results of triglycerides conc. in the present investigation are harmonized with that reported by Le *et al.* (2016) for the suckling Holstein calves. Likewise, Talha *et al.* (2009) and Noori *et al.* (2016) indicated that cholesterol values were significantly ( $P < 0.05$ ) reduced by the probiotics treatment, which agreed with our findings.

After 15 days of the probiotics treatment end, the significant rise of serum IgG conc. in G2 and G3 are similar to that noticed by Chen *et al.* (2021) who recorded that probiotics (*B. subtilis*, *B. licheniformis* and *Lactobacillus plantarum*) treatment increased the conc. of IgG in lambs. In the same trend, beef calves supplemented with *B. amyloliquefaciens*/*B. subtilis*, the serum IgG levels were increased ( $P > 0.05$ ). Serum IgG are produced by B-lymphocytes, which are the major impact factors of humoral immunity, to prevent and resist infection (Du *et al.*, 2018). Moreover, there was a tendency for increasing the IgG1 conc. with the probiotics (*B. subtilis* + *B. licheniformis*) addition in milk replacer on the 45<sup>th</sup> day of life of suckling Holstein calves (Riddell *et al.*, 2010). Supplementing *Bacillus* based probiotics to the diet would stimulate an increase in IgG1 levels as an anti-spore immune response (Hong *et al.*, 2005).

### Effect of Probiotics Supplements on Health Status

The lower dose of probiotics supplementation (G2) showed a lower incidence of diarrhea (ID)

and duration of diarrhea (DD) and the incidence of pneumonia (IP) compared with the control (G1) and the higher dose of probiotics supplementation (Figs. 2 and 3). These results are consistent with no mortality problems; also no respiratory problems were detected all over the experimental period.

These results are similar to those obtained by Mousa and Marwan (2019) who found that pathogenic microbes number was decreased and the beneficial microbes number was increased which reflected in low diarrhea incidence. Also, Agazzi *et al.* (2014) and Kowalski *et al.* (2009) reported that using lactic acid bacteria in suckling animals lowered the incidence and frequency of diarrhea and showed slightly lower fecal score. In contrast, many authors noticed a significant reduction in health problem incidences when calves received the probiotics (Abe *et al.*, 1995; Abu-Tarboush *et al.*, 1996; Timmerman *et al.*, 2005) which may be related to the reaction of calves on probiotics depends on the conditions in which the experiment is being conducted.

The differences between some previous studies and results in this study might be due to the feeding strategy, environmental conditions, diet composition, type and dose of the supplemented probiotics.

### Economic Feed Efficiency

The economic evaluation was based on the current selling price of the tested diets (liquid and dry), the tested additives and the kilograms of live body weight had shown in (Table 5).

The price per each Kg of milk, starter mixture, probiotic and weight gain were 7.50, 4, 24, 240 and 127,5 E£, respectively. The relative profit = (average daily profit of G1, G2 and G3/ average daily profit of G1) × 100.

The results showed that the adding the lower dose of probiotics (10g/head/day) increased the total return/head/day 120.70% compare with the total return of the control one (100%). However, adding the higher dose of probiotics (20 g/head/day) increased the total return/head/day only by 101.22% compared with the control diet. There was no discernible difference between the higher supplemented dose and the control.



Fig. 2. Effects of probiotics supplementation on percentages of diarrhea incidence (ID %) and diarrhea duration (DD, in days)

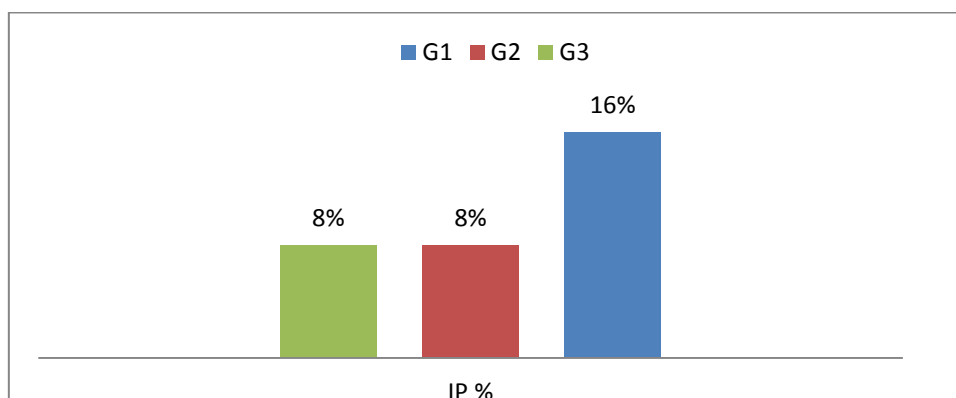


Fig. 3. Effects of probiotics supplementation on percentages of incidence of pneumonia (IP %) before weaning

Table 5. Effect of probiotics supplementation on the economic feed efficiency of the suckling Holstein calves.

Items	Groups			
	G <sub>1</sub>	G <sub>2</sub>	G <sub>3</sub>	
<b>Total consumption:</b>				
Milk	Kg/h	510	510	510
Starter mixture	Kg/h	18.60	21.58	18.93
<b>Total cost:</b>				
Milk	E£	3825	3825	3825
Starter mixture	E£	78.86	89.04	81.40
Probiotics	E£	0.00	72	144
Total d cost	E£	3903.86	3986.04	4050.40
<b>Average daily cost</b>	<b>E£</b>	65.06	66.43	67.50
<b>Total gain:</b>				
Total gain	Kg/ 60 day	39.16	47.25	39.62
Average daily gain	Kg/day	0.652	0.787	0.660
<b>Average daily gain</b>	<b>E£</b>	71.72	86.57	72.60
<b>Daily gain %</b>	<b>E£</b>	100	120.70	101.22
<b>Average daily profit</b>	<b>E£</b>	6.66	20.27	5.10
<b>Relative profit</b>	<b>%</b>	100	304.35	76.58

## Conclusion

It can be concluded that the performance and health status of the suckling Holstein calves were improved by using 10 g/head/day of *Bacillus strains (subtillis and licheniformis)* spores as safe alternatives to antibiotics.

## REFERENCES

- Abe, F., N. Ishibashi and S. Shimamura (1995). Effect of administration of bifidobacteria and lactic acid bacteria to newborn calves and piglets. *J. Dairy Sci.*, 78 (12): 2838-2846.
- Abu-Tarboush, H.M., M.Y. Al-Saiady and A.H.K. El-Din (1996). Evaluation of diet containing lactobacilli on performance, fecal coliform, and lactobacilli of young dairy calves. *Anim. Feed Sci. and Technol.*, 57 (1-2): 39-49.
- Afsharmanesh, M. and B. Sadaghi (2014). Effects of dietary alternatives (probiotics, green tea powder, and Kombucha tea) as antimicrobial growth promoters on growth, ileal nutrient digestibility, blood parameters, and immune response of broiler chickens. *Comparative Clin. Pathol.*, 23 (3): 717-724.
- Agazzi, A., E. Tirloni, S. Stella, S. Marocolo, B. Ripamonti, C. Bersani, J. Caputo, V. Dell'Orto, N. Rota and G. Savoini (2014). Effects of species-specific probiotics addition to milk replacer on calf health and performance during the first month of life. *Ann. Anim. Sci.*, 14(1): 101-115.
- Aikman, P.C., P.H. Henning, D.J. Humphries, and C.H. Horn (2011). Rumen pH and fermentation characteristics in dairy cows supplemented with *Megasphaera elsdenii* NCIMB 41125 in early lactation. *J. Dairy Sci.*, 94 (6): 2840-2849.
- Amburgh, M.Van. and J.A.M.E.S. Drackley (2005). Current perspectives on the energy and protein requirements of the pre-weaned calf. Calf and heifer rearing: principles of rearing the modern dairy heifer from calf to calving. 60<sup>th</sup> Univ. Nottingham Easter School in Agric. Sci., Nottingham, UK. 23<sup>rd</sup>-24<sup>th</sup> March, 2004, 67-82.
- An, Y., S.P. Young, S.L. Hillman, J.L. Van Hove, Y.T. Chen and D.S. Millington (2000). Liquid chromatographic assay for a glucose tetra saccharide, a putative biomarker for the diagnosis of Pompe disease. *Anal. Biochem.*, 287 (1): 136-143.
- AOACO (1990). Official Methods of Analysis. Association of Official Analytical Chemists, Arlington, VA.
- Arora, D.S. and N. Kaur (2020). Prospecting the antimicrobial and antibiofilm potential of *Chaetomium globosum* an endophytic fungus from *Moringa oleifera*. *AMB Express*, 10 (1): 206.
- Bai, S.P., A.M. Wu, X.M. Ding, Y. Lei, J. Bai, K.Y. Zhang and J.S. Chio (2013). Effects of probiotics-supplemented diets on growth performance and intestinal immune characteristics of broiler chickens. *Poult. Sci.*, 92 (3): 663-670.
- Bakker, A.J. and M. Mücke (2007). Gammopathy interference in clinical chemistry assays mechanisms, detection and prevention.
- Boyd, J., J.W. West and J.K. Bernard (2011). Effects of the addition of direct-fed microbials and glycerol to the diet of lactating dairy cows on milk yield and apparent efficiency of yield. *J. Dairy Sci.*, 94 (9): 4616-4622.
- Chen, J., O.M. Harstad, T. McAllister, P. Dörsch and H. Holo (2020). Propionic acid bacteria enhance ruminal feed degradation and reduce methane production in vitro. *Acta Agric. Scandinavica, Section A-Anim. Sc.*, 69 (3): 169-175.
- Chen, H., B. Guo, M. Yang, J. Luo, Y. Hu, M. Qu and X. Song (2021). Response of growth performance, blood biochemistry indices, and rumen bacterial diversity in lambs to diets containing supplemental probiotics and chinese medicine polysaccharides. *Frontiers in Vet. Sci.*, 656.
- Duncan, D.B. (1955). Multiple range and multiple F- tests. *Biomet.*, 11(1): 1-42.
- Du, R., S. Jiao, Y. Dai, J. An, J. Lv, X. Yan and B. Han (2018). Probiotics *Bacillus amyloliquefaciens* C-1 improves growth

- performance, stimulates GH/IGF-1, and regulates the gut microbiota of growth-retarded beef calves. *Frontiers in Microbiol.*, 9: 2006.
- Engvall, E. and P. Perlmann (1971). Enzyme-linked immunosorbent assay (ELISA) quantitative assay of immunoglobulin G. *Immunochem.*, 8 (9): 871-874.
- FAO/WHO (2002). Guidelines for the Evaluation of probiotics in Food. *Food and Agriculture Organization of the United Nations/World Health Organization*, London, Ontario.
- Fouladgar, S., A.F. Shahraki, G.R. Ghalamkari, M. Khani, F. Ahmadi and P.S. Erickson (2016). Performance of Holstein calves fed whole milk with or without kefir. *J. Dairy Sci.*, 99 (10): 8081-8089.
- Hong, H.A., L.H. Duc and S.M. Cutting (2005). The use of bacterial spore formers as probiotics. *FEMS Microbiol. Rev.*, 29 (4): 813-835.
- Hussein, H.A., J.P. Thurmann and R. Staufenbiel (2020). 24-h variations of blood serum metabolites in high yielding dairy cows and calves. *BMC Vet. Res.*, 16 (1): 1-11.
- Kochewad, S.A., A.B. Kanduri, J.M. Chahande, D.S. Deshmukh, S.A. Ali and V.M. Patil (2009). Effect of probiotics supplementation on Growth parameters of Osmanabadi Kids.
- Khalifa, E.I., H.A. Hassanien, A.H. Mohamed, A.M. Hussein and A.A. Abd-Elaal (2016). Influence of addition *Spirulina platensis* algae powder on reproductive and productive performance of dairy Zaraibi goats. *Egypt. J. Nutr. and Feeds*, 19 (2): 211-225.
- Kowalski, Z.M., P. Górka, A. Schlagheck, W. Jagusiak, P. Micek and J. Strzetelski (2009). Performance of Holstein calves fed milk-replacer and starter mixture supplemented with probiotics feed additive. *J. Anim. and Feed Sci.*, 18 (3): 399-411.
- Layus, B.I., C.L. Gerez and A.V. Rodriguez (2020). Antibacterial activity of *Lactobacillus plantarum* CRL 759 against methicillin-resistant *Staphylococcus aureus* and *Pseudomonas aeruginosa*. *Arab. J. Sci. and Eng.*, 45 (6): 4503-4510.
- Le, O.T., P.J. Dart, K. Harper, D. Zhang, B. Schofield, M.J. Callaghan and D.M. McNeill (2016). Effect of probiotics *Bacillus amyloliquefaciens* strain H57 on productivity and the incidence of diarrhoea in dairy calves. *Anim. Prod. Sci.*, 57(5): 912-919.
- Liao, Q., X. Hang, X. Liu, J. Pan, H. Zhang and H. Yang (2010). The influence of pH on heat stress response by probiotics *Lactobacillus plantarum* LP-Onlly. *Ann. Microbiol.*, 60 (2): 341-348.
- Ma, Z.Z., Y.Y. Cheng, S.Q. Wang, J.Z. Ge, H.P. Shi and J.C. Kou (2020). Positive effects of dietary supplementation of three probiotics on milk yield, milk composition and intestinal flora in Sannan dairy goats varied in kind of probiotics. *J. Anim. Physiol. and Anim. Nutr.*, 104 (1): 44-55.
- Maas, R.M., M.C. Verdegem, S. Debnath, L. Marchal and J.W. Schrama (2021). Effect of enzymes (phytase and xylanase), probiotics (*B. amyloliquefaciens*) and their combination on growth performance and nutrient utilisation in Nile tilapia. *Aquac.*, 533: 736226.
- Mookiah, S., C.C. Siew, K. Ramasamy, N. Abdullaha and Y.W. Hoa (2014). Effects of dietary prebiotics, probiotics and synbiotics on performance, caecal bacterial populations and caecal fermentation concentrations of broiler chickens. *J. Sci. Food and Agric.*, 94: 341-348.
- Mostafa, T.H., F.A. Elsayed, M.A. Ahmed and M.A. Elkholy (2014). Effect of using some feed additives (tw-probiotics) in dairy cow rations on production and reproductive performance. *Egypt. J. Anim. Prod.*, 51 (1): 1-11.
- Mousa, S.A. and A.A. Marwan (2019). Growth performance, rumen fermentation and selected biochemical indices in buffalo calves fed on *Basillus subtilis* supplemented diet. *Int. J. Vet. Sci.*, 8 (3): 151-156.
- Mousa, S., A. Elsayed, B. Marghani and A. Ateya (2019). Effects of supplementation of *Bacillus* spp. on blood metabolites, antioxidant status, and gene expression pattern of selective cytokines in growing Barki lambs. *J. Adv. Vet. Anim. Res.*, 6 (3): 333-340.

- Muya, M.C., F.V. Nherera, K.A. Miller, C.C. Aperce, P.M. Moshidi and L.J. Erasmus (2015). Effect of *Megasphaera elsdenii* NCIMB 41125 dosing on rumen development, volatile fatty acid production and blood  $\beta$ -hydroxybutyrate in neonatal dairy calves. *J. Anim. Physiol. and Anim. Nutr.*, 99 (5): 913-918.
- National Research Council (2001). Nutrient Requirements of dairy cattle: 2001. National Academies Press.
- Noori, M., M. Alikhani and R. Jahanian (2016). Effect of partial substitution of milk with probiotics yogurt of different pH on performance, body conformation and blood biochemical parameters of Holstein calves. *J. Appl. Anim. Res.*, 44 (1): 221-229.
- Park, J.H., S.J. Cho and J.M. Cox-Ganser (2016). Observational scores of dampness and mold associated with measurements of microbial agents and moisture in three public schools. *Indoor Air*, 26 (2): 168-178.
- Renaud, D.L.L., B.J.N. Wilms and M.A. Steele (2020). Technical note: Is fecal consistency scoring an accurate measure of fecal dry matter in dairy calves? *J. Dairy Sci.*, 103: 10709–10714.
- Riddell, J.B., A.J. Gallegos, D.L. Harmon and K.R. Mcleod (2010). Addition of a *Bacillus* based probiotics to the diet of preruminant calves: Influence on growth, health, and blood parameters<sup>1, 2, 3</sup>. *Int. J. Appl. Res. Vet. M.*, 8: 78-85.
- Rifai, N., P.M. Ridker, M.A. Pfeffer, F. Sacks, and E. Braunwald (1999). Long-term effects of pravastatin on plasma concentration of C-reactive protein. *The Cholesterol and Recurrent Events (CARE) Investigators. Circulation*, 100 (3): 230–235.
- Seifzadeh, S., F. Mirzaei Aghjehgheshlagh, H. Abdibenemar, J. Seifdavati and B. Navidshad (2017). The effects of a medical plant mix and probiotics on performance and health status of suckling Holstein calves. *Italian J. Anim. Sci.*, 16 (1): 44-51.
- Sharma, S., S. Kandasamy, D. Kavitate and P.H. Shetty (2018). Probiotics characterization and antioxidant properties of *Weissella confusa* KR780676, isolated from an Indian fermented food. *LWT*, 97: 53-60.
- Signorini, M.L., L.P. Soto, M.V. Zbrun, G.J. Sequeira, M.R. Rosmini and L.S. Frizzo (2012). Impact of probiotics administration on the health and fecal microbiota of young calves: a meta-analysis of randomized controlled trials of lactic acid bacteria. *Res. Vet. Sci.*, 93(1), 250-258.
- Smock, T.M., K.L. Samuelson, J.E. Hergenreder, P.W. Rounds and J.T. Richeson (2020). Effects of *Bacillus subtilis* PB6 and/or chromium propionate supplementation on clinical health, growth performance, and carcass traits of high-risk cattle during the feedlot receiving and finishing periods. *Translational Anim. Sci.*, 4 (3): txaa163.
- Talha, M.H., R.I. Moawd, A.A. Abu El-Ella and G.H. Zaza (2009). Effect of some feed additives on rearing calves from birth till weaning: 1–Productive performance and some blood parameters. *J. Anim. and Poultry Prod.*, 34 (4): 2763-2783.
- Thomas, L. (1998). Alanine aminotransferase (ALT), Aspartate aminotransferase (AST). *Clinical Laboratory Diagnostics*. 1<sup>st</sup> Ed. Frankfurt: TH-Books Verlagsgesellschaft, 2: 55-65.
- Timmerman, H.M., L. Mulder, H. Everts, D.C. van Espen, E. van der Wal, G. Klaassen, S.M. Rouwers, R. Hartemink, F.M. Rombouts and A.C. Beynen (2005). Health and growth of veal calves fed milk replacers with or without probiotics. *J. Dairy Sci.*, 88 (6): 2154–2165.
- Van Zyl, W.F., S.M. Deane and L.M. Dicks (2020). Molecular insights into probiotics mechanisms of action employed against intestinal pathogenic bacteria. *Gut Microbes*, 12 (1): 1831339.
- Zhang, Y., Gu, Y., Ren, H., Wang, S., Zhong, H., Zhao, X. ... and Wang, W. (2020). Gut microbiome-related effects of berberine and probiotics on type 2 diabetes (the PREMOTE study). *Nature Communications*, 11 (1):1-12.
- Zhang, Z.F. and I.H. Kim (2014). Effects of multistrain probiotics on growth performance, apparent ileal nutrient digestibility, blood characteristics, caecal microbial shedding, and excreta odor contents in broilers. *Poultry Sci.*, 93 (2): 364-370.

## أثر استخدام البروبيوتيك المتكون من (*Bacillus subtilis* و *Bacillus licheniformis*) على الأداء الحيوي وصحة العجول الهولشتاين الرضيعة

أحمد السيد شتاوي – صبري محمد بسيوني – أسامة محمد عبدالمنعم – خالد محمد المرابي

قسم الإنتاج الحيواني – كلية الزراعة – جامعة الزقازيق - مصر

كان الهدف من هذه الدراسة هو تقييم تأثير إضافة جراثيم *Bacillus subtilis* و *Bacillus licheniformis* على إجمالي المادة الجافة المأكولة ومعدل النمو اليومي والاستفادة من العلف وعمر الفطام وحالات حدوث الإسهال ومدته وحالات حدوث الالتهاب الرئوي وبعض مكونات مصل الدم في عجول الهولشتاين الرضيعة. تم تقسيم عدد 36 من عجول الهولشتاين النقية حديثة الولادة (عمر 5 أيام، ومتوسط وزن  $44,27 \pm 0,37$  و  $42,87 \pm 0,55$  كجم للذكور والإناث، على الترتيب) بشكل عشوائي إلى ثلاث مجموعات متجانسة (12 عجلاً/مجموعة، 6 عجول من كل جنس). تم تغذية جميع المجموعات على اللبن الكامل ثلاث مرات يومياً لمدة 30 يوماً متصلة بواسطة دلاء الرضاعة. وتمت إذابة صفر و10 و20 جرام من مسحوق البروبيوتيك في اللبن كامل الدسم لتمثل G1 (المجموعة الضابطة) وG2 وG3 على التوالي. ويحتوي كل جرام من البروبيوتيك على  $2 \times 10^9$  من جراثيم *Bacillus subtilis* وكذلك  $2 \times 10^9$  من جراثيم *Bacillus licheniformis*. بدأ تقديم مخلوط العلف اليومي للعجول في اليوم الثامن من العمر. وتمت دراسة نفس المعالم المختبرة لجميع مجموعات العجول لمدة شهر آخر بعد انتهاء معاملة البروبيوتيك. أظهرت النتائج المتحصل عليها أن قيمة إجمالي المادة الجافة المأكولة لم تتغير أثناء فترة المعاملة بالبروبيوتيك (30 يوماً)، لكنها زادت بشكل ملحوظ ( $P < 0.001$ ) خلال الشهر التالي للمعاملة في G2 عن تلك الموجودة في G1 وG3. وخلال مرحلتي التجربة، كان معدل النمو اليومي للعجول في G2 أفضل ( $P < 0.001$ ) من ذلك في المجموعات المختبرة الأخرى. وفي نهاية التجربة، كانت عجول G2 أثقل وزناً بمقدار 8,68 كجم من عجول المجموعة الضابطة. كانت جميع قياسات كفاءة الاستفادة من الغذاء أفضل ( $P < 0.001$ ) مع الجرعة الأقل من البروبيوتيك (G2). وقد ترتب على ذلك أن كان عمر الفطام في G2 أبكر منه في بقية المجموعات المختبرة. وقد انخفض معدل حدوث الإسهال ومدته في عجول G2 مقارنة بالعجول في G1 وG3، بينما لوحظت أعلى نسبة حدوث للالتهاب الرئوي مع عجول G1 وذلك خلال كامل فترة التجربة. ولم يكن هناك اختلاف معنوي ( $P \geq 0.05$ ) لتركيزات الجلوكوز والبروتين الكلي والكرياتينين ولأنشطة ALT وAST في سيرم الدم عجول المجموعات المختبرة عند نهاية المعاملة بالبروبيوتيك. ومن ناحية أخرى فقد انخفضت مستويات الألبومين ( $P = 0.02$ ) في G2 وG3، بينما ارتفع مستوى الجلوبيولين ( $P = 0.02$ ) في G2 مقارنة مع G1 ولكنها ظلت في معدلاتها الطبيعية. بالإضافة إلى ذلك، كانت قيم الدهون الثلاثية والكوليسترول أقل بشكل معنوي ( $P = 0.001$ ) في G2 وG3 من تلك الموجودة في G1. في حين أنه قد ارتفعت معنوياً ( $P = 0.001$ ) تركيزات الجلوبيولين المناعي IgG في مصل الدم في G2 وG3 بعد 15 يوماً من نهاية المعاملة بالبروبيوتيك مقارنة بتلك الموجودة في G1. وقد أظهر التقييم الاقتصادي لتغذية عجول الهولشتاين الرضيعة عائداً أفضل مع المجموعة التي تلقت الجرعة الأقل من البروبيوتيك (G1) مقارنة بالمجموعات الأخرى. ويمكن أن نجمل بأن إضافة جراثيم *Bacillus subtilis* + *Bacillus licheniformis* إلى اللبن كامل الدسم كانت لها آثاراً مفيدة على أداء وصحة عجول الهولشتاين الرضيعة.

### المحكمون:

رئيس بحوث بمعهد بحوث الإنتاج الحيواني – مركز البحوث الزراعية.  
أستاذ تغذية الحيوان المساعد – كلية الزراعة – جامعة الزقازيق.

1- أ.د. أيمن عبدالحى عبدالحاميد  
2- د. أدهم عبدالله الصغير